Updates with respect to the previous version are marked in grey.

Anti-SARS-CoV-2 ELISA (IgG)

Instruction for use

For in vitro diagnostic use IVD

ORDER NO.	ANTIBODIES AGAINST	IG CLASS	SUBSTRATE	FORMAT
EI 2606-9601 G EI 2606-9620 G	SARS-coronavirus-2 (SARS-CoV-2)	IgG	Ag-coated microplate wells	96 x 01 (96) 96 x 20 (1920)

CE

Intended use

The enzyme immunoassay (ELISA) provides semiquantitative in vitro determination of human antibodies of the immunoglobulin class IgG against SARS-CoV-2 in serum, EDTA, heparin or citrate plasma or dried blood spots (DBS) to support the diagnosis of SARS-CoV-2 infection and constitutes a supplement to the direct pathogen detection. Moreover, serology can be applied to collect epidemiological data. The product is designed for use as IVD and can optionally be processed on fully automated equipment. The format 96 x 20 has been specially designed for processing on the EUROLabWorkstation ELISA.

Clinical significance

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2, previously called 2019-nCoV) belongs to the family of coronaviruses and, like SARS-CoV, is classified in the genus *Betacoronavirus* [1]. At the end of 2019, SARS-CoV-2 was identified as the causative agent of clustered cases of pneumonia of unclear origin. The virus caused an infection wave which quickly spread worldwide and was declared a pandemic by the WHO at the beginning of 2020 [2-5].

SARS-CoV-2 is predominantly transmitted by droplet infection via coughing or sneezing and through close contact with infected persons [3-4, 6]. Healthcare personnel and family members are especially at risk of infection [6]. The zoonotic reservoir of the virus appears to be bats [3, 4, 6].

The incubation time of SARS-CoV-2 is three to seven, maximally 14 days [2]. The symptoms of SARS-CoV-2 infection are fever, coughing, breathing difficulties and fatigue [2-4, 6]. In most patients the infection manifests with symptoms of a mild febrile illness with irregular lung infiltrates. Some patients, especially elderly or chronically ill patients, develop acute respiratory distress syndrome (ARDS) [2, 3, 5, 6]. In February 2020, the disease caused by SARS-CoV-2 was named COVID-19 by the WHO.

Suitable methods for the diagnosis of SARS-CoV-2 infections are detection of viral RNA by reverse transcriptase polymerase chain reaction (RT-PCR) or of viral protein by ELISA primarily in sample material from the upper (nasopharyngeal or oropharyngeal smear) or lower respiratory tract (bronchoalveolar lavage fluid, tracheal secretion, sputum, etc.). The detection of viral antigens is less sensitive than RT-PCR.

The determination of antibodies enables confirmation of SARS-CoV-2 infection in patients with typical symptoms and in suspected cases. It also contributes to monitoring and outbreak control [4, 5]. The spike (S) and nucleocapsid (N) proteins of SARS-CoV-2 are highly immunogenic. Antibodies against receptor binding domain (RBD) of the spike protein are neutralising [7]. For significant serological results, two patient samples should be investigated, one from the acute phase (week 1 of the illness) and one from the convalescent phase (3 to 4 weeks later) [4, 6, 8]. Cross-reactions with antibodies within the genus *Betacoronavirus* have been described [4, 5].

The majority of COVID-19 patients in the convalescent phase show a pronounced T-cell reactivity to SARS-CoV-2. Cross-reacting T-cells are found in around half of the patients without SARS-CoV-2 history. This is presumably due to past infections with coronaviruses causing common colds. SARS-CoV-2-specific T-cells are part of the T-cell repertoire of persons who had an infection with SARS-CoV in 2003. These cells proliferate following contact with SARS-CoV-2. These findings indicate a long-term immunity after infection with Betacoronaviruses [9].

Antigen

The reagent wells of the ELISA were coated with an S1 domain of the spike protein of SARS-CoV-2 expressed recombinantly in the human cell line HEK 293.

Test principle

The test kit contains microplate strips each with 8 break-off reagent wells coated with recombinant S1 domain of the spike protein of SARS-CoV-2. The format EI 2606-9620 G has been optimised for processing with the EUROLabWorkstation ELISA and contains components in the EUROTank. Information on automated incubation is given in the instructions for use of the respective instruments. In the first reaction step, diluted patient samples are incubated in the wells. In the case of positive samples, specific IgG (also IgA and IgM) antibodies will bind to the antigens. To detect the bound antibodies, a second incubation is carried out using an enzyme-labelled anti-human IgG (enzyme conjugate) catalysing a colour reaction.

Contents of the test kit

Component	Colour	Format 96 x 01	Format 96 x 20	Symbol
1. Microplate wells coated with antigens 12 microplate strips each containing 8 individual break-off wells in a frame, ready for use	-	12 x 8	12 x 8 x 20	STRIPS
2. Calibrator (IgG, human), ready for use	dark red	1 x 2.0 ml	4 x 2.0 ml	CAL
3. Positive control (IgG, human), ready for use	blue	1 x 2.0 ml	4 x 2.0 ml	POS CONTROL
4. Negative control (IgG, human), ready for use	green	1 x 2.0 ml	4 x 2.0 ml	NEG CONTROL
5. Enzyme conjugate peroxidase-labelled anti-human IgG, ready for use	green	1 x 12 ml	3 x 80 ml	CONJUGATE
6. Sample buffer ready for use	light blue	1 x 100 ml	11 x 100 ml	SAMPLE BUFFER
7. Wash buffer 10x concentrate	colourless	1 x 100 ml	10 x 100 ml	WASH BUFFER 10x
8. Chromogen/substrate solution TMB/H_2O_2, ready for use	colourless	1 x 12 ml	3 x 80 ml	SUBSTRATE
9. Stop solution 0.5 M sulphuric acid, ready for use	colourless	1 x 12 ml	3 x 80 ml	STOP SOLUTION
10. EUROTank cap, black	-	-	5 pieces	CAP
11. Protective foil	-	3 pieces	-	FOIL
12. Quality control certificate	-	1 protocol	1 protocol	-
13. Test instruction	-	1 booklet	-	-

Additional materials and equipment (not supplied in the test kit)

- EUROLabWorkstation ELISA (EUROIMMUN order no. YG 0851-0101) only for format 9620
- Automatic microplate washer: recommended. Washing of the microplates can also be carried out manually.
- Microplate reader: wavelength of 450 nm, reference wavelength range from 620 nm to 650 nm
- Calibrated pipettes
- Pipette tips
- Stepper pipette: recommended for the pipetting of enzyme conjugate, substrate, and stop solution
- Distilled or deionised water
- Incubator: for incubation of the microplate at +37°C
- Incubator or water bath: recommended to warm the wash buffer
- Stop watch

Depending on the instrument, further materials are required for automatic processing. For more information, please refer to the respective instructions for use.

Storage and stability

The test kit has to be stored at a temperature between +2°C and +8°C, do not freeze. Unopened, all test kit components are stable until the indicated expiry date.

In use stability following the first opening

After opening, the reagents are stable until the indicated expiry date when stored at +2°C to +8°C and protected from contamination, unless stated otherwise below.

Warnings and precautions

- The product must only be used by trained laboratory personnel in a clinical or research laboratory.
- If the packed reagents are visibly damaged, do not use the test kit.
- Before using the product, read the instruction for use carefully. Only the valid version is to be used. This is provided with the test kit or can be obtained from the customer portal (https://products.euroimmun.de).
- Do not substitute or mix the EUROIMMUN reagents with reagents from other manufacturers.
- Observe Good Laboratory Practice (GLP) and safety guidelines. Some of the reagents contain preservatives in non-declarable concentrations. Avoid eye and skin contact with samples and reagents. In case of eye or skin contact, rinse thoroughly with water. Remove and wash contaminated clothing. In case of ingestion, obtain medical advice.
- The calibrator and controls of human origin have tested negative for HBsAg, anti-HCV, anti-HIV-1 and anti-HIV-2. Nonetheless, all reagents should be treated as being a potential infection hazard and should be handled with care.

Preparation and stability of the samples

- **Samples:** Human serum or EDTA, heparin or citrate plasma or dried capillary blood (dried blood spots, DBS), collected with the EUROIMMUN Blood Collection Card (EUROIMMUN order number ZV 9711-01100).
- **Sample preparation: Patient samples** are diluted **1:101** in sample buffer.

  For example: dilute 10 µl serum in 1.0 ml sample buffer and mix well by vortexing (sample pipettes are not suitable for mixing).

  Note: When using instruments for automated incubation, sample handling is described in the instructions for use.

  When dried blood spots (DBS) are used as the sample material, these must be extracted from the membrane of the blood collection card prior to sample incubation. The test instruction required for the extraction (EUROIMMUN document number EI_2606G_A_UK_ZXX) is provided in the customer portal (https://products.euroimmun.de).
- **Stability of the patient samples:**
  - stored at +2°C to +8°C: up to 14 days
  - incubate diluted samples within one working day

Preparation and stability of the reagents

Note: All reagents must be brought to room temperature (+18°C to +25°C) approx. 30 minutes before use.

The thermostatically adjustable ELISA incubator must be set to +37°C ± 1°C.

- **Coated wells:** Ready for use. Tear open the resealable protective wrapping of the microplate at the recesses above the grip seam. Do not open until the microplate has reached room temperature to prevent the strips from moistening. Immediately replace the remaining wells of a partly used microplate in the protective wrapping and tightly seal with the integrated grip seam (Do not remove the desiccant bag).
  Once the protective wrapping has been opened for the first time, the wells coated with antigens can be stored in a dry place and at a temperature between +2°C and +8°C for 4 months.
- **Calibrator and controls:** Ready for use. Mix reagents thoroughly before use.
- **Enzyme conjugate:** Ready for use. Mix the reagent thoroughly before use.
- **Sample buffer:** Ready for use.
- **Wash buffer:** The wash buffer is a 10x concentrate. If crystallisation occurs in the concentrated buffer, warm it to +37°C and mix well before dilution. Dilute the required volume 1:10 with deionised or distilled water (1 part reagent plus 9 parts water).
  For example: For 1 microplate strip, 5 ml concentrate plus 45 ml water.
  The working-strength wash buffer is stable for 4 weeks if stored at +2°C to +8°C and handled properly.
- **Chromogen/substrate solution:** Ready for use. Close the tube immediately after use, as the contents are sensitive to light. The chromogen/substrate solution must be clear on use. Do not use the solution if it is blue coloured.
- **Stop solution:** Ready to use.

Waste disposal

Patient samples, calibrators, controls and incubated microplate strips should be handled as infectious waste. All reagents must be disposed of in accordance with local disposal regulations.

Quality control

For every group of tests performed, the extinction readings of the calibrator and ratios determined for the positive and negative controls must lie within the limits stated for the relevant test kit lot. A quality control certificate containing these reference values is included. If the values specified for the controls are not achieved, the test results may be inaccurate and the test should be repeated.

Reference material

As no quantified international reference serum exists for antibodies against SARS-CoV-2, the calibration is performed in ratios which are a relative measure for the concentration of antibodies in serum or plasma.

Assay procedure

(Partly) manual test performance

ATTENTION: The processing of the format EI 2606-9620 G on the EUROLabWorkstation ELISA is based on the assay stored there. For details, please refer to the instructions for use of the instrument.

Sample incubation: (1st step)
Transfer **100 µl** of the **calibrator, positive and negative controls or diluted patient samples** into the individual microplate wells according to the pipetting protocol. Incubate for **60 minutes** at **+37°C ± 1°C**.
For manual processing of microplate wells, cover the finished test plate with the protective foil. When using an automated microplate processor for incubation, follow the recommendations of the instrument manufacturer.

Washing:
Manual: Remove the protective foil. Empty the wells and subsequently wash **3 times using 300 µl of working-strength wash buffer** for each wash.
Automatic: Remove the protective foil. Wash the reagent wells **3 times with 450 µl of working-strength wash buffer** (program setting: e.g. TECAN Columbus Washer “Overflow Mode”).

Leave the wash buffer in each well for 30 to 60 seconds per washing cycle, then empty the wells. After washing (manual and automated tests), thoroughly dispose of all liquid from the microplate by tapping it on absorbent paper with the openings facing downwards to remove all residual wash buffer.

Note:
Free positions on the microplate strip should be filled with blank wells of the same plate format as that of the parameter to be investigated.

Conjugate incubation: (2nd step)
Pipette **100 µl of enzyme conjugate** (peroxidase-labelled anti-human IgG) into each of the microplate wells. For manual test performance cover the reagent wells with the protective foil.
Incubate **30 minutes at +37°C ± 1°C**.

Washing:
Remove the protective foil. Empty the wells. Wash as described above.

Substrate incubation: (3rd step)
Pipette **100 µl of chromogen/substrate solution** into each of the microplate wells. Incubate for **30 minutes** at room temperature (+18°C to +25°C) protected from direct sunlight.

Stopping:
Pipette **100 µl of stop solution** into each of the microplate wells in the same order and at the same speed as the chromogen/substrate solution was introduced.

Measurement:
Photometric measurement of the colour intensity should be made at a **wavelength of 450 nm** and a reference wavelength between 620 nm and 650 nm **within 30 minutes of adding the stop solution.** Prior to measuring, slightly shake the microplate to ensure a homogeneous distribution of the solution.

Test performance using fully automated analysis devices

Sample dilution and test performance are carried out fully automatically using an analysis device. The incubation conditions programmed in the respective software authorised by EUROIMMUN may deviate slightly from the specifications given in the ELISA test instruction, but have been validated in respect of the combination of the EUROIMMUN Analyzer I, the EUROIMMUN Analyzer I-2P, the EUROLabWorkstation ELISA, the Sprinter XL and the DSX from Dynex and this EUROIMMUN ELISA Validation documents are available on enquiry.

Note: Processing on other fully automated systems is possible but must be validated by the user.

Pipetting protocol

	1	2	3	4	5	6	7	8	9	10	11	12
A	C	P 6	P 14	P 22								
B	pos.	P 7	P 15	P 23								
C	neg.	P 8	P 16	P 24								
D	P 1	P 9	P 17									
E	P 2	P 10	P 18									
F	P 3	P 11	P 19									
G	P 4	P 12	P 20									
H	P 5	P 13	P 21									

The pipetting protocol for microplate strips 1 to 4 is an example for the **semiquantitative analysis** of 24 patient sera (P 1 to P 24).

The calibrator (C), the positive (pos.) and negative (neg.) controls, and the patient samples have each been incubated in one well. The reliability of the ELISA test can be improved by duplicate determinations for each sample.

The wells can be broken off individually from the strips. This makes it possible to adjust the number of test substrates used to the number of samples to be examined and minimises reagent wastage.

Both positive and negative controls serve as internal controls for the reliability of the test procedure. They should be assayed with each test run.

Test evaluation

ATTENTION: The test evaluation for the format EI 2606-9620 G is performed with the software of the EUROLabWorkstation ELISA. For details, please refer to the instructions for use of the instrument.

The extinction of the calibrator defines the upper limit of the reference range of non-infected persons (**cut-off**) recommended by EUROIMMUN. Values above the indicated cut-off are to be considered as positive, those below as negative.

Semiquantitative: Results can be evaluated semiquantitatively by calculating a ratio of the extinction of the control or patient sample over the extinction of the calibrator. Calculate the ratio according to the following formula:

$$\frac{\text{Extinction of the control or patient sample}}{\text{Extinction of calibrator}} = \text{Ratio}$$

EUROIMMUN recommends interpreting results as follows:

Ratio <0.8: Negative
Ratio ≥ 0.8 to <1.1: Borderline
Ratio ≥1.1: Positive

For duplicate determinations the mean of the two values should be taken. If the two values deviate substantially from one another, EUROIMMUN recommends retesting the samples.

Analytical performance

The following data were collected using serum or plasma samples:

Measurement range:

Limit of blank (LoB): ratio 0.13
Limit of detection (LoD): ratio 0.14

LoB and LoD were defined according to the requirements defined in guideline EP17-A2 of the CLSI (Clinical and Laboratory Standards Institute, https://clsi.org/).

Precision: Studies on the intra-lab precision were carried out according to CLSI guideline EP05-A3. Six samples (reactivity distributed over the entire measurement range) were measured. The precision is given as standard deviation (SD) and coefficient of variation (CV).

Intra-lab precision

	Sample 1		Sample 2		Sample 3		Sample 4		Sample 5		Sample 6	
Mean	Ratio 4.74		Ratio 2.74		Ratio 1.09		Ratio 1.25		Ratio 1.14		Ratio 0.22	
	SD	% CV	SD	% CV	SD	% CV	SD	% CV	SD	% CV	SD	% CV
Repeatability	0.125	2.6	0.071	2.6	0.032	2.9	0.064	5.1	0.028	2.5	0.028	12.6
Between run	0.173	3.6	0.172	6.3	0.067	6.1	0.089	7.1	0.115	10.1	0.023	10.5
Within day	0.213	4.5	0.186	6.8	0.074	6.8	0.109	8.7	0.119	10.4	0.036	16.4
Between day	0.139	2.9	0.000	0.0	0.028	2.5	0.047	3.8	0.000	0.0	0.025	11.3
Within lab	0.254	5.4	0.186	6.8	0.079	7.3	0.119	9.5	0.119	10.4	0.044	19.9

Cross-reactivity (analytical specificity): Due to low homologies of the S1 protein within the coronavirus family, cross-reactions to most of the human pathogenic representatives of this virus family are virtually excluded. However, due to their close relationship, cross-reactions between SARS-CoV(-1) and SARS-CoV-2 are likely. Sera from patients with SARS-CoV(-1), MERS-CoV, HCoV-229E, HCoV-NL63, HCoV-HKU1 or HCoV-OC43 infections were investigated to examine this further. As expected, there were pronounced cross-reactivities, especially with anti-SARS-CoV(-1) IgG antibodies. Cross-reactions to other human pathogenic coronaviruses were not observed.

Interference: Haemolytic, lipaemic and icteric samples showed no influence on the result up to concentrations of 10 mg/ml haemoglobin, 20 mg/ml triglycerides and 0.4 mg/ml bilirubin in this ELISA.

The following data were collected using DBS samples:

Precision: Studies on the intra-lab precision were carried out according to CLSI guideline EP05-A3. Six samples (reactivity distributed over the entire measurement range) were measured. The data were determined on 15 days in two runs per day with two replicates each. Each replicate was yielded by independent extraction from a dried blood spot. The precision is given as standard deviation (SD) and coefficient of variation (CV).

Intra-lab precision

	Sample 1		Sample 2		Sample 3		Sample 4		Sample 5		Sample 6	
Mean	Ratio 0.26		Ratio 0.90		Ratio 0.99		Ratio 1.38		Ratio 3.73		Ratio 5.95	
	SD	% CV	SD	% CV	SD	% CV	SD	% CV	SD	% CV	SD	% CV
Repeatability	0.045	17.0	0.114	12.6	0.069	7.0	0.140	10.1	0.251	6.7	0.377	6.3
Between run	0.000	0.0	0.000	0.0	0.115	11.6	0.083	6.1	0.292	7.8	0.000	0.0
Within day	0.045	17.0	0.114	12.6	0.134	13.5	0.163	11.8	0.385	10.3	0.377	6.3
Between day	0.025	9.6	0.028	3.1	0.051	5.2	0.053	3.9	0.000	0.0	0.141	2.4
Within lab	0.051	19.5	0.117	12.9	0.143	14.5	0.171	12.4	0.385	10.3	0.402	6.8

Method comparison:

Study I: The correlation between extracts of dried blood spots (DBS) from capillary blood and serum from venous blood was determined by analysing 215 patient samples collected in Europe, using the Anti-SARS-CoV-2 ELISA (IgG). For each patient, one capillary blood sample and one venous blood sample were available.
The agreement between the results of the dried capillary blood spots and the venous blood samples was 100% (positive agreement (PPA): 100%; negative agreement (NPA): 100%). Borderline samples were excluded from the calculation.

n = 215		**EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) Sera from venous blood**		
		positive	**borderline**	**negative**
EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) DBS	**positive**	61	2	0
	borderline	0	1	4
	negative	0	0	147

Study II: In another study, 51 patient samples (origin: Brazil) were investigated using the Anti-SARS-CoV-2 ELISA (IgG). Like in the first study, extracts from dried blood spots and serum samples from venous blood were compared.
The agreement between the results of the dried capillary blood spots and the venous blood samples was 100% (positive agreement (PPA): 100%; negative agreement (NPA): 100%). Borderline samples were excluded from the calculation.

n = 51		**EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) Sera from venous blood**		
		positive	**borderline**	**negative**
EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) DBS	**positive**	40	0	0
	borderline	0	1	0
	negative	0	0	10

Clinical performance

Diagnostic sensitivity (Prevalence):

To determine the diagnostic sensitivity, samples from patients with confirmed SARS-CoV-2 infection were analysed. The following sensitivity therefore corresponds to the prevalence of antibodies against SARS-CoV-2 in COVID-19 infected persons.

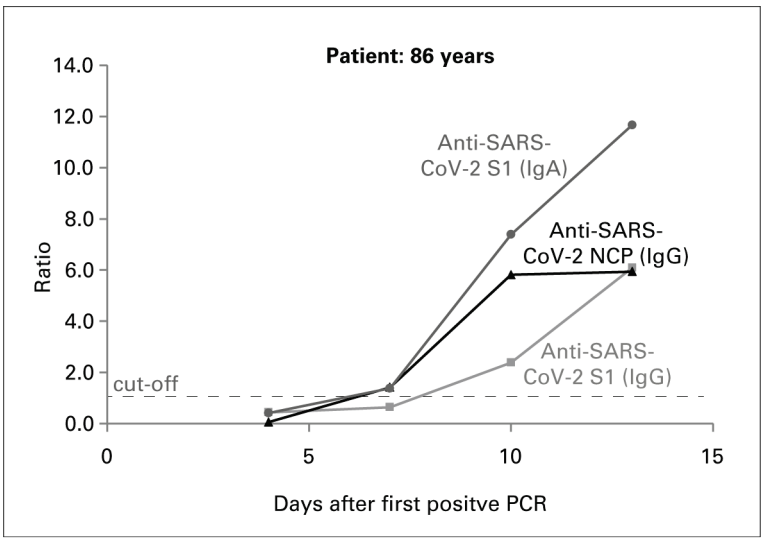
The sensitivity was determined by investigating 166 samples from 152 European patients, using the Anti-SARS-CoV-2 ELISA (IgG). In these patients, infections with SARS-CoV-2 had been confirmed by RT-PCR [10] based on a sample taken at the early phase of infection. In samples taken prior to day 10 (time point after onset of symptoms or positive direct pathogen detection), the Anti-SARS-CoV-2 ELISA (IgG) showed a sensitivity of 43.7%. The sensitivity of the Anti-SARS-CoV-2 ELISA (IgG) in samples collected after day 10 was 94.4%. Borderline results (n = 7) were not considered in the calculation.

Days after symptom onset or positive direct pathogen detection	EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG)		
	Positive	Negative	Sensitivity
≤ 10	38	49	**43.7%**
> 10	68	4	**94.4%**

The time course of antibody formation and the antibody activity at specific time points can vary significantly. In most patients, antibodies are detectable after day 10 after symptom onset or positive direct pathogen detection. In individual cases, a significantly delayed synthesis of IgG (> 4 weeks after onset of symptoms or positive direct pathogen detection) has been reported. The graphs show individual immune responses in COVID-19 patients measured with the EUROIMMUN Anti-SARS-CoV-2 ELISA (IgA) and the EUROIMMUN Anti-SARS-CoV-2 ELISA (IgG) based on the recombinant S1 domain of the spike protein, and the EUROIMMUN Anti-SARS-CoV-2 NCP ELISA (IgG), for which a modified nucleocapsid protein (NCP) is used as antigen.

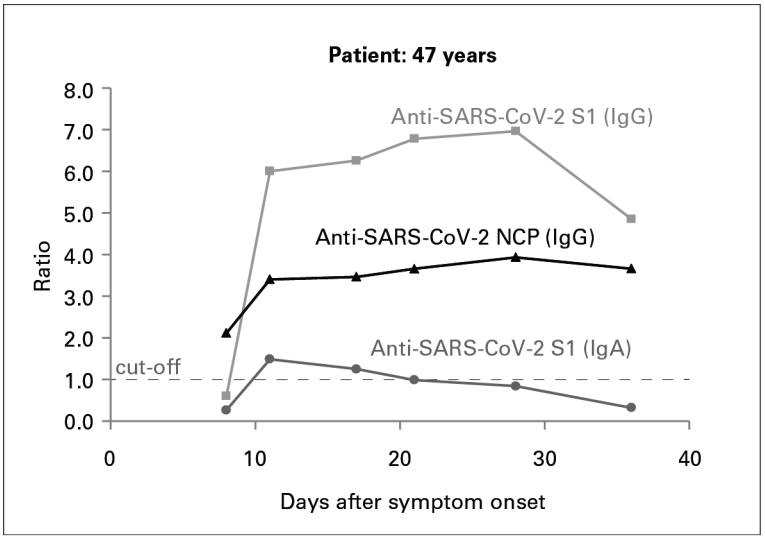
Patient 1 (86 years old)

Anti-SARS-CoV-2 S1 IgA and anti-SARS-CoV-2 NCP IgG antibodies were detectable as early as 7 days after RT-PCR. The level of anti-SARS-CoV-2 S1 IgG antibodies was still negative 7 days after positive RT-PCR, but was increased in the subsequent sample taken on day 10.



Patient 2 (47 years old)

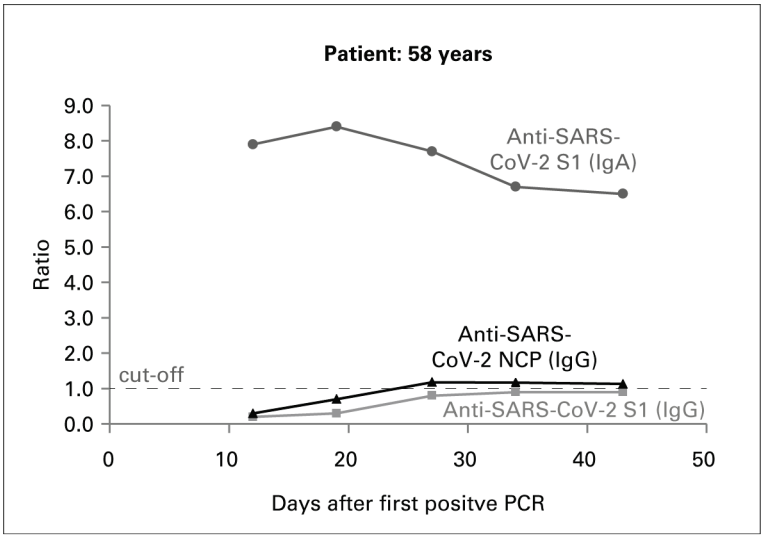
The anti-SARS-CoV-2 NCP IgG antibody level was elevated as early as day 8 after the onset of symptoms. Anti-SARS-CoV-2 S1 IgA and IgG antibodies were not yet detectable. A follow-up sample taken on day 11 after the onset of symptoms showed an increase in the antibody levels for both Ig classes.



Patient 3 (58 years old)

The anti-SARS-CoV-2 S1 IgA antibody level was already highly elevated 12 days after positive RT-PCR. In contrast, the levels of anti-SARS-CoV-2 S1 and anti-SARS-CoV-2 NCP IgG antibodies increased only slowly until day 43 after positive RT-PCR.

Note: In individual cases, delayed antibody formation may occur, so that antibodies are only detectable after a period of several weeks after the onset of symptoms.



Specificity: The specificity of the Anti-SARS-CoV-2 ELISA (IgG) was determined by analysing 222 patient samples that were positive, for instance, for antibodies against other human pathogenic coronaviruses, other pathogens or for rheumatoid factors. Additionally, 1122 samples from blood donors, children and pregnant women obtained before the occurrence of SARS-CoV-2 (before January 2020) were analysed. The results in the borderline range (n = 7) were not considered in the calculation. The specificity of the Anti-SARS-CoV-2 ELISA (IgG) amounted to 99.6%.

Panel	**n**	**EUROIMMUN Anti-SARS-CoV-2 ELISA IgG**
		Specificity
Blood donors	849	99.5%
Pregnant women	199	99.5%
Children	74	100.0%
Elderly people	97	100.0%
Infections with other human pathogenic coronaviruses	23	100.0%
Influenza (freshly vaccinated, including courses)	40	100.0%
Acute EBV infections & heterophile antibodies	22	100.0%
Rheumatoid factors	40	100.0%
Total	**1344**	**99.6%**

Limitations of the procedure

- For a medical diagnosis, the serological test result should always be interpreted together with the clinical symptoms of the patient and other results, e.g. those of the direct pathogen detection.
- A negative serological result does not exclude an infection. Particularly in the early phase of infection, antibodies may not yet be present or are only present in such small quantities that they are not detectable. In the case of a borderline result, a secure evaluation is not possible. If there is a clinical suspicion and a negative or borderline test result, we recommend clarification by means of other diagnostic methods and/or the serological investigation of a follow-up sample. A positive result indicates that there has been contact with the pathogen. In the determination of pathogen-specific IgM antibodies, polyclonal stimulation of the immune system or antibody persistence may affect the diagnostic relevance of positive findings. Significantly higher specific IgG antibody levels (increase by more than factor 2) and/or seroconversion in a follow-up sample taken after at least 7 to 10 days can indicate an acute infection. Sample and follow-up sample should be incubated in parallel in adjacent wells of the ELISA microplate within the same test run.
- The pipetting volumes, incubation times, temperatures, and preparation steps given in the instruction for use must be adhered to.
- Correct performance of sample collection and storage is crucial for the test results.
- The test system is validated for the determination of anti-SARS-CoV-2 IgG in human serum, plasma or dried blood spots only.
- The binding activity of the antibodies and the activity of the enzyme used are temperature-dependent. It is therefore recommended using a thermostatically adjusted ELISA incubator in all incubation steps. The higher the room temperature during the incubation steps, the greater will be the extinction. The same variations also apply to the incubation times. However, the calibrators are subject to the same influences, with the result that such variations will be largely compensated in the calculation of the result.

- Insufficient washing (e.g. less than 3 wash cycles, too small wash buffer volumes, or too short residence times) can lead to false high extinction readings.
- Residual liquid (>10 µl) in the reagent wells after washing can interfere with the substrate and lead to false low extinction readings.
- The partial or complete adjustment of the test system to the use of instruments for automated sample processing or other liquid handling devices may result in differences between the results obtained with automated processing and those obtained with manual procedure. It is the responsibility of the user to validate the instruments used so that they yield test result within the reliable range.

Literature

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Technical support

In case of technical problems you can obtain assistance via the EUROIMMUN website (www.euroimmun.com/contact).

Meaning of the symbols

Symbol	Meaning	Symbol	Meaning
STRIPS	Microplate strips	LOT	Lot description
CAL	Calibrator		Protect from sunlight
POS CONTROL	Positive control		Storage temperature
NEG CONTROL	Negative control		Unopened usable until (YYYY-MM-DD)
CONJUGATE	Conjugate	CE	CE-labelled
SAMPLE BUFFER	Sample buffer		Manufacturing date (YYYY-MM-DD)
WASH BUFFER 10x	Wash buffer, 10x concentrate		Manufacturer
SUBSTRATE	Substrate		Observe instructions for use
STOP SOLUTION	Stop solution	REF	Order number
FOIL	Protective foil		Contents suffice for <n> analyses
CAP	Cap		Biological risks
IVD	In vitro diagnostic medical device		